

FORUM REVIEW ARTICLE

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# A Hibernation-Like State for Transplantable Organs: Is Hydrogen Sulfide Therapy the Future of Organ Preservation?

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## Abstract

**Significance:** Renal transplantation is the treatment of choice for end-stage renal disease, during which renal grafts from deceased donors are routinely cold stored to suppress metabolic demand and thereby limit ischemic injury. However, prolonged cold storage, followed by reperfusion, induces extensive tissue damage termed cold ischemia/reperfusion injury (IRI) and puts the graft at risk of both early and late rejection.

**Recent Advances:** Deep hibernators constitute a natural model of coping with cold IRI as they regularly alternate between 4°C and 37°C. Recently, endogenous hydrogen sulfide (H<sub>2</sub>S), a gas with a characteristic rotten egg smell, has been implicated in organ protection in hibernation.

**Critical Issues:** In renal transplantation, H<sub>2</sub>S also seems to confer cytoprotection by lowering metabolism, thereby creating a hibernation-like environment, and increasing preservation time while allowing cellular processes of preservation of homeostasis and tissue remodeling to take place, thus increasing renal graft survival.

**Future Directions:** Although the underlying cellular and molecular mechanisms of organ protection during hibernation have not been fully explored, mammalian hibernation may offer a great clinical promise to safely cold store and reperfuse donor organs. In this review, we first discuss mammalian hibernation as a natural model of cold organ preservation with reference to the kidney and highlight the involvement of H<sub>2</sub>S during hibernation. Next, we present recent developments on the protective effects and mechanisms of exogenous and endogenous H<sub>2</sub>S in preclinical models of transplant IRI and evaluate the potential of H<sub>2</sub>S therapy in organ preservation as great promise for renal transplant recipients in the future. *Antioxid. Redox Signal.* 00, 000–000.

**Keywords:** renal transplantation, ischemia-reperfusion injury, hibernation, graft function, organ preservation, hydrogen sulfide (H<sub>2</sub>S)

## Introduction

RENAL TRANSPLANTATION is the preferred therapeutic option for end-stage renal disease. Compared with dialysis, kidney transplantation offers the best long-term outcomes in terms of survival, quality of life, and cost-effectiveness (23). Renal grafts may be derived from living donors or deceased

(i.e., brain dead or nonheart beating) donors and are routinely flushed with and stored in cold preservation solutions such as University of Wisconsin (UW) solution at 4°C to decrease energy demand during this storage period. Hypothermic kidney storage before transplantation results in ischemic kidney damage due to restriction of blood flow to the kidney. The final stage of ischemic injury occurs during reperfusion, the effector

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phase of ischemic injury, which develops when blood flow is restored in the graft. The whole process is defined as ischemia/reperfusion injury (IRI). Cold storage preserves graft quality, thus allowing time for transportation and recipient selection, compared with warm IRI (10). However, the beneficial effects of cold storage of donor kidneys are limited. While short-term cold storage reduces cellular oxygen demand and thereby prevents tissue injury, prolonged cold storage leads to renal injury. The ensuing cold ischemic injury is characterized by tubular injury and epithelial cell death, thereby increasing the incidence of delayed graft function (DGF) and lowering graft survival (27). It has been reported that every 6-h increase in cold storage increases the risk of DGF by 23% (64). Furthermore, prolonged cold storage is associated with worse long-term graft survival (68). Thus, cold IRI is unavoidable, but remains the major contributor to early and late graft dysfunction following renal transplantation.

The mechanisms underlying cold IRI are complex and not fully elucidated. Some studies have reported that such events include mitochondrial injury evidenced by depletion of mitochondrial adenosine triphosphate (ATP) leading to loss of cellular homeostasis and thereby affecting several cellular pathways involved in tissue regeneration and repair (68, 78). Furthermore, excess production of free radicals in the mitochondria seems to play a key role in cold-induced mitochondrial injury (59) and may lead to inflammation and apoptosis of glomerular and tubular cells of cold-stored kidneys (55). The restoration of blood supply and temperature in renal graft during reperfusion results in excessive reactive oxygen species (ROS) generation (21, 55), which initiates a cascade of events, including activation of inflammatory and apoptotic pathways, leading to tissue injury (57). Unfortunately, the search for therapeutic strategies to mitigate cold IRI has met little success so far. Thus, the quest for safe reduction of metabolism in transplant organs continues.

It is of interest to note that mammalian hibernators are able to survive periods of low metabolic rate and body temperature without signs of renal injury in contrast to nonhibernating species. Hibernators alternate their body temperature between 4°C and 37°C without cold IRI challenge. Recently, hydrogen sulfide (H<sub>2</sub>S), a gas with a distinctive smell of rotten eggs and the third member of the gasotransmitter family, has been implicated in organ protection in deep hibernation (80) and also seems to confer cytoprotection by lowering metabolism and increasing preservation time and overall increasing renal graft survival after transplantation (2, 30, 33, 34). H<sub>2</sub>S is produced enzymatically in all mammalian species, including humans (52). Two of these enzymatic pathways are cytosolically catalyzed by cystathionine β-synthase (CBS) and cystathionine γ-lyase (CSE). The third enzymatic pathway requires the mitochondrial enzyme, 3-mercaptopyruvate sulfurtransferase (3-MST), while the fourth pathway involves the peroxisomal enzyme, D-amino acid oxidase (DAO) (74, 89).

Apart from being produced endogenously, there are various forms of exogenous H<sub>2</sub>S that have been used experimentally in the form of donor molecules. The classic form of inhalation is seriously hampered by toxic effects to bystanders, while sulfide salts (sodium hydrosulfide [NaHS] and Na<sub>2</sub>S) are short-lasting and may not always reach their target sites, particularly the mitochondria (45). Fortunately, slow-releasing H<sub>2</sub>S donors such as GYY4137 (51) and mitochondria-targeting slow-releasing H<sub>2</sub>S donors (AP39

and AP123) have been developed (35, 49, 79) and may enable translation of fundamental findings on H<sub>2</sub>S in modulating renal IRI into human kidney preservation.

In this review, we first discuss mammalian hibernation as a unique natural model of how nature deals with cold IRI with reference to the kidney as a typical organ highly vulnerable to cold IRI. We also highlight the protective role of H<sub>2</sub>S during hibernation. Next, we present recent findings on H<sub>2</sub>S as a gasotransmitter that confers cytoprotection during cold IRI in renal transplantation, suggesting that it may become a novel method of preservation to help improve graft quality and increase the long-term success of renal transplantation in the future.

### Lessons Learned from Mammalian Hibernation

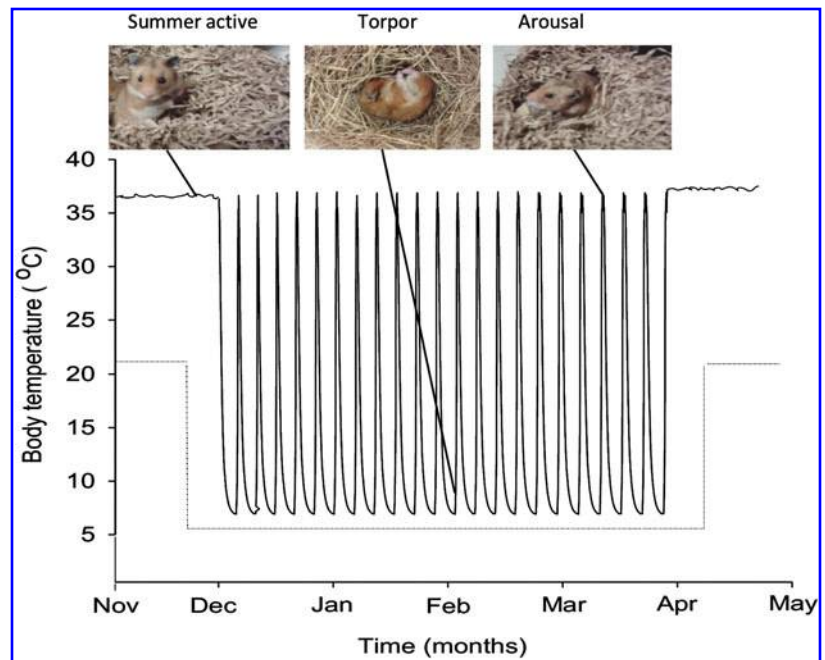
Mammalian hibernation may offer a great clinical promise to safely cold store and reperfuse donor organs. Hibernation is an evolved strategy that conserves energy mainly seasonally and confers a significant survival advantage to individuals among select, but diverse, mammalian lineages (67). To save energy demand during winter, mammalian hibernators undergo repetitive cycles of torpor and arousal (12, 26) (Fig. 1). Torpor is a state of reduced metabolism and body temperature usually between 4°C and 10°C, depending on the species (13), and can be viewed as the natural equivalent of cold storage of donor organs. As illustrated in Figure 1, the torpor phase lasting days to weeks is intermittently interrupted by brief periods of arousal during which metabolism and body temperature return to euthermic levels (40). Hence, the arousal phase may represent a natural equivalent of reperfusion after cold storage.

While the body temperature of smaller mammalian hibernators falls drastically during torpor and lasts days to weeks, followed by intermittent arousals, larger hibernators such as bears remain in torpid state at 30–35°C for 5–7 months in a year without eating coupled with decreased metabolism, immobilization, and anuria due to complete reabsorption of the glomerular filtrate (17, 76, 84) (Fig. 2). A more extreme type of torpor is seen in hibernating Arctic ground squirrels, during which core body temperature measures –2.9°C without freezing and metabolism reduced to 1% of euthermic rate (5). Interestingly, whereas torpor and arousal cause no organ injury in all hibernating animals studied thus far (19, 42, 43, 51, 92), cold storage and reperfusion result in extensive organ injury in nonhibernating species (30, 42), including humans.

In the kidney, for example, the morphology of the renal cortex and medulla is not affected by hibernation in typical hibernating animals such as 13-lined ground squirrels and dormice, which are the most studied hibernating animals (43, 70, 92). Moreover, the ultrastructure of the glomerular basement membrane, tubular brush border, and cytoskeleton are well preserved throughout torpor and arousal—a condition that would induce extensive damage in human deceased donor kidneys (43, 92). Renal protection in hibernating animals is such a strong intrinsic event that kidneys removed from hibernating 13-lined ground squirrels and stored in cold UW solution resisted apoptotic cell death as shown by reduced caspase-3 activity (42, 43).

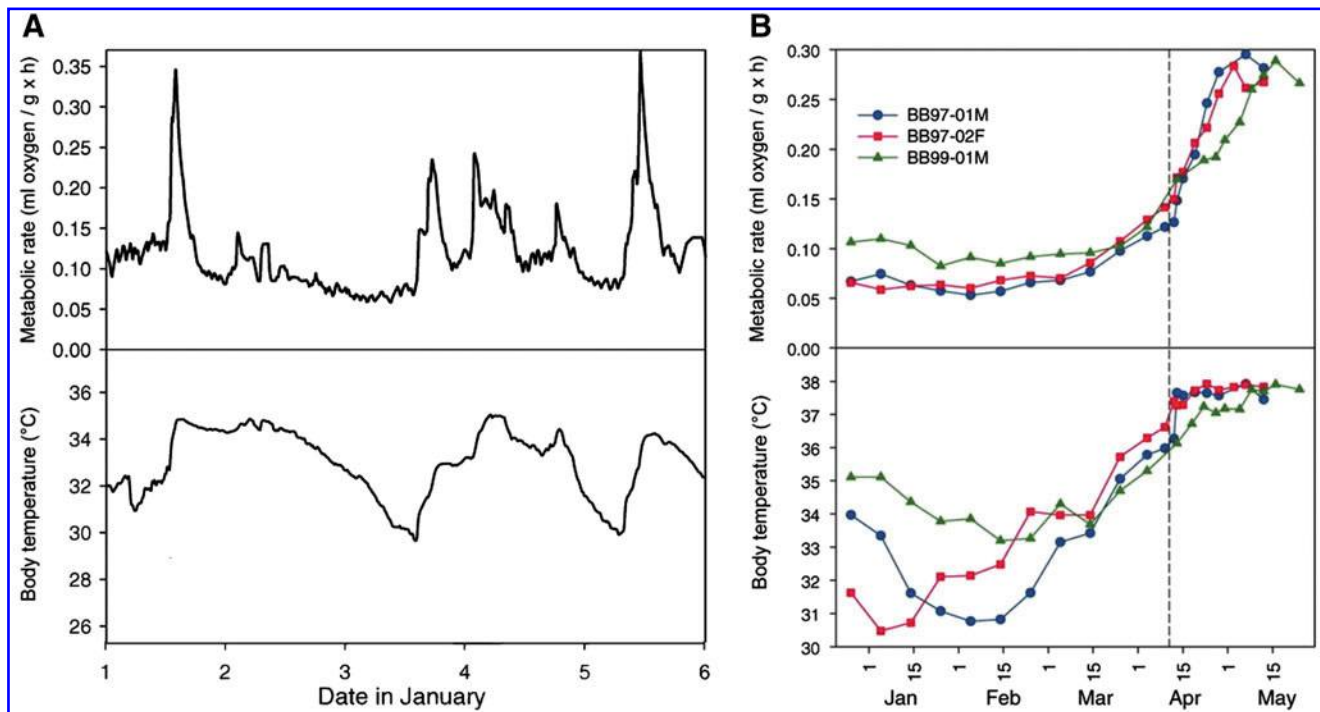
The activation and suppression of mitochondrial metabolism and organ protection in hibernating animals seem tightly regulated, and the molecular mechanism of this regulation is not fully understood. Although it is yet to be identified which

**FIG. 1. Torpor–arousal cycle of a Syrian golden hamster with respect to appearance and core Tb.** The graph shows a simulation of Tb tracings (solid line) of a Syrian hamster (*Mesocricetus auratus*) inside a climate-controlled chamber. The dashed line represents the ambient temperature, which is lowered from 21°C to 5°C to induce hibernation and back to 21°C to end the torpor–arousal cycle. Tb dropped from ~37°C to ~7°C as ambient temperature was lowered. Periodic arousals between each torpor bout are associated with restoration of euthermic temperature of ~37°C despite constant ambient temperature. Photographs of hamsters at three different time points during the hibernation are shown above the graph. Tb, body temperature.



endogenous molecules are essential for entrance into torpor and/or arousal, H<sub>2</sub>S has so far proven important in the torpor–arousal cycle as its level is markedly increased in the lung during torpor and reduced in the arousal state in Syrian hamsters (80). This finding suggests that H<sub>2</sub>S may influence temperature-controlling neurotransmitters such as histamine, serotonin, and opioids, which have been proposed to play key

roles in regulation of the torpor pattern by the central nervous system (38, 47, 69) as well as phospholipids of liver cell nuclei (48). Additionally, organ protection in hibernation has been attributed to increased production of endogenous H<sub>2</sub>S, which can also induce a torpor-like state in small nonhibernating mammals with effects on mitochondrial metabolism (8, 9, 80) and endogenous antioxidants such as glutathione and



**FIG. 2. Core body temperature and metabolic rate during torpor and arousal in bears.** (A) Body temperature and oxygen consumption of a black bear (*Ursus americanus*) housed in an outside den during a 5-day period in January. (B) Body temperature and oxygen consumption of three bears during torpor and arousal. Vertical dashed line represents average time of emergence, which varied by  $\pm 2$  days. Data are adapted from a study by Tøien *et al.* (84).

ascorbic acid (28), as well as upregulation of antiapoptotic genes such as B cell lymphoma-2 (Bcl-2) and phospho-Akt (31, 42). The renoprotective effect of H<sub>2</sub>S was also observed in rats (nonhibernating animals), in which dopamine treatment upregulated renal expression of CBS, CSE, and 3-MST and stimulated endogenous H<sub>2</sub>S production, leading to protection against cold renal IRI (30). Dopamine or serotonin treatment of rat smooth muscle aortic cells as well as rat kidney and other tissues before cold storage also protected against hypothermia/rewarming-induced apoptosis and subsequent hypothermic injury by strongly increasing the endogenous H<sub>2</sub>S level (81).

In addition, our group demonstrated that endogenous H<sub>2</sub>S production protects kidneys of hamsters (natural hibernators) subjected to 5'-adenosine monophosphate (AMP)-induced torpor-like state at 7°C body temperature (29) (Figs. 3 and 4). Furthermore, Blackstone and Roth demonstrated that pretreatment with inhaled H<sub>2</sub>S for only 20 min markedly prolonged survival without any apparent detrimental effects in mice exposed to otherwise lethal hypoxia (5% oxygen) (9). It appears that kidneys and tissues of mammalian hibernators are well protected from the damaging effects of hypothermia and reperfusion through upregulated antioxidant pathways and maintenance of mitochondrial homeostasis. These findings suggest that H<sub>2</sub>S plays an important role in modulating cytoprotective pathways that govern protection against cold IRI and allow maintenance of mitochondrial function throughout torpor and arousal.

### Do Organs of Hibernators Suffer Ischemia During Torpor or Is There an Element of Organ Preservation?

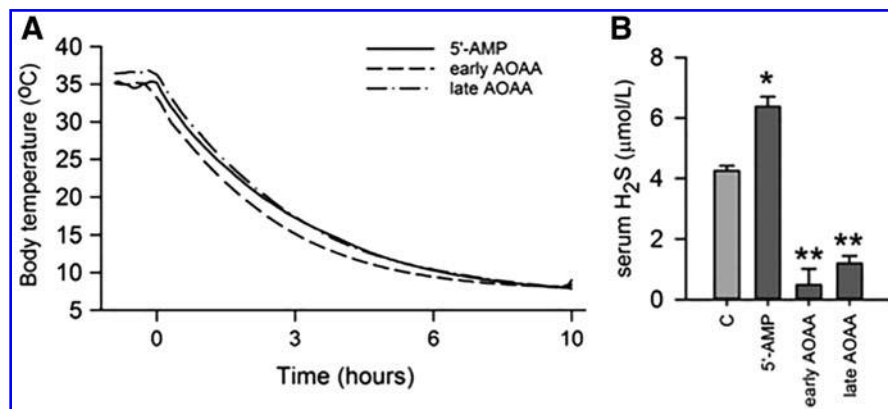
Since torpor is characterized by reduced metabolism and body temperature, a major question that immediately comes to mind is whether the profound reduction in cardiac output and oxygenation of the kidney and other tissues during hibernation results in renal ischemia during hibernation. The kidneys of nonhibernating animals in a healthy state receive about 20% of cardiac output, which mostly goes to the renal cortex, while the renal medulla receives only about 10% of

the total renal perfusion (7), implying that the renal medulla is highly vulnerable to ischemia even at euthermic temperature. From a clinical point of view, ischemia occurs when there is an imbalance between blood oxygen demand and supply, which may result in cell death and tissue injury (15). To date, there are no good data to strongly suggest whether clinical renal ischemia occurs during torpor and, if it occurs, whether it may serve as a primary stimulus for arousal from torpor. Likely, the profound lowering of metabolism during torpor runs parallel with the reduced perfusion, which ensures a balance between blood oxygen demand and supply and thereby prevents ischemia during torpor (22).

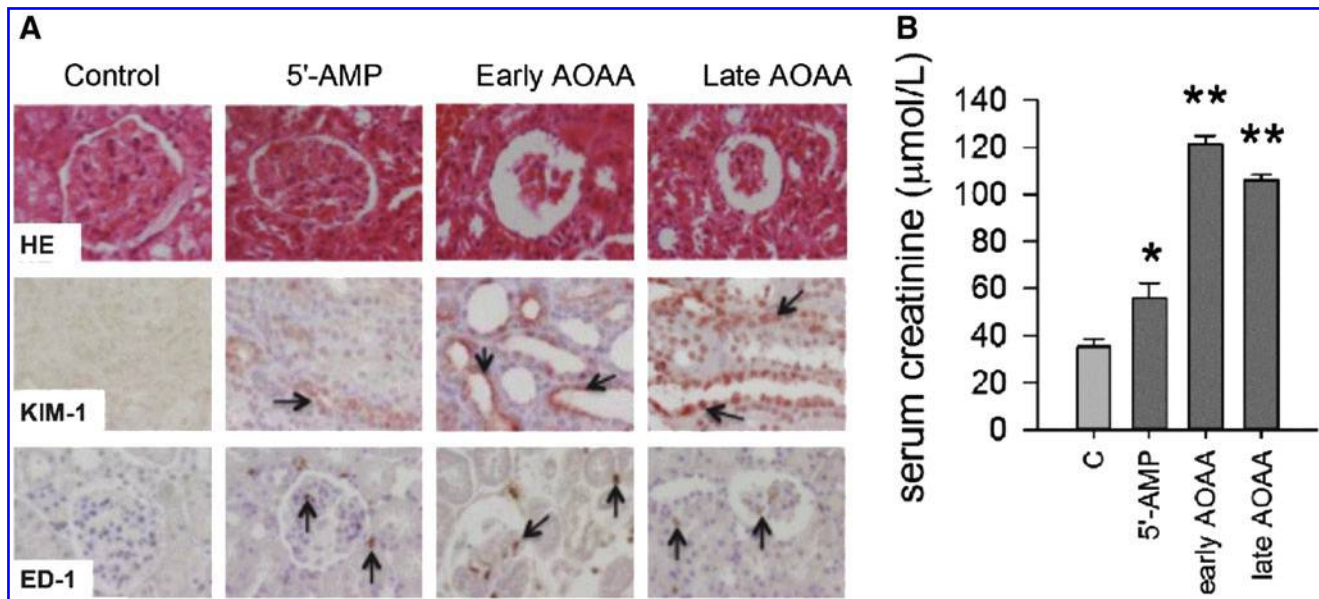
Despite the debate whether or not torpor is associated with (renal) ischemia/reperfusion (I/R), the profound increase in metabolic rate upon rewarming after torpor likely results in high levels of oxidative stress and can potentially lead to cellular injury if not counterbalanced by protective mechanisms. Specific adaptations, consisting of upregulation of antioxidant enzymes and antiapoptotic pathways, limit oxidative stress and protect against cell death throughout hibernation (14). It should be noted that in contrast to hypothermic I/R, arousal from torpor may not be associated with a gross fluctuation in tissue oxygenation. In the brain of torpid Arctic ground squirrels, for example, there is a high arterial oxygen pressure and low expression of hypoxia inducible factor- $\alpha$ , a marker of tissue hypoxia (56), suggesting that hibernators may not show signs of ischemia during torpor. In conclusion, it remains to be revealed whether hibernators suffer renal ischemia from a clinical perspective during torpor and arousal.

### H<sub>2</sub>S Lowers Metabolic Rate at High Concentrations, but Stimulates ATP Production at Low Concentrations

At high concentration of 80 ppm, for example, H<sub>2</sub>S can suppress metabolism through reversible inhibition of mitochondrial complex IV (cytochrome c oxidase) (6, 8). Thus, it is not surprising that H<sub>2</sub>S affects metabolism in animals. Inhibition of mitochondrial respiration was implicated in the induction of suspended animation in mice (6). Inhalation



**FIG. 3.** 5'-AMP induces torpor in natural hibernators and increases serum H<sub>2</sub>S level. (A) 5'-AMP administration (at t = 0) resulted in a drop in body temperature from 37°C to ~7°C following 10 h of 5'-AMP injection, which was not affected by early or late administration of AOAA. (B) 5'-AMP administration markedly increased the plasma H<sub>2</sub>S level compared with control animals, while administration of AOAA before or 4 h after 5'-AMP injection reduced the serum H<sub>2</sub>S level. \*\*/\**p* < 0.05/0.01 compared with control. Data are presented as mean ± SEM. AMP, adenosine monophosphate; AOAA, amino-oxycetic acid; C, control animals; H<sub>2</sub>S, hydrogen sulfide; SEM, standard error of the mean. Data are adapted from study by Dugbartey *et al.* (29).



**FIG. 4. Blocking endogenous H<sub>2</sub>S production during 5'-AMP-induced torpor induces renal injury.** (A) Representative photographs of kidney tissue from immuno(histochemical) staining (magnification  $\times 400$ ). Pharmacological inhibition of endogenous H<sub>2</sub>S during 5'-AMP-induced torpor provokes kidney damage characterized by glomerular atrophy, tubular dilatation, atrophy of epithelial cells, and widening of tubular lumen. (B) Serum creatinine level showing the degree of renal functional derangement following pharmacological inhibition of endogenous H<sub>2</sub>S in 5'-AMP-induced torpor. Arrows point to positively stained areas (in brown), indicating tubular injury and inflammation. ED-1, antibody against CD68 specific for macrophages; HE, hematoxylin and eosin; KIM-1, Kidney Injury Molecule 1. Data are adapted from study by Dugbartey *et al.* (29).

of gaseous H<sub>2</sub>S induces a hibernation-like state in non-hibernating mice by profoundly reducing the metabolic rate and core body temperature and protecting against cold renal IRI (8, 9, 11). Inhibition of oxidative phosphorylation by reversible antagonism at complex IV of the mitochondrial respiratory chain has been proposed to underlie the induction of such a hypometabolic state by H<sub>2</sub>S (8).

Whereas high dosages cause hypometabolism, low concentrations of H<sub>2</sub>S (0.1–1  $\mu$ M) enhance electron transport in the mitochondria by acting as an electron donor at the level of coenzyme Q—an electron carrier between complexes I and III in the mitochondrial electron transport chain (ETC). Such electron donation by H<sub>2</sub>S stimulates cellular energy metabolism and ATP production through oxidative phosphorylation (36, 60). Recent studies also suggest that in the low-oxygen environment of ischemia/hypoxia, CBS and CSE enzymes translocate from the cytosol to the mitochondria to augment mitochondrial H<sub>2</sub>S production by 3-MST through electron donation and thus protecting liver and heart mitochondria against ischemic injury (32, 83). Perhaps the mitochondrial protection and preservation of ATP synthesis offered by H<sub>2</sub>S at low concentrations explain why hibernators are protected from cold ischemic injury. In conclusion, H<sub>2</sub>S at high concentration lowers metabolism, but at low concentration acts as an electron donor and preserves mitochondrial function.

#### H<sub>2</sub>S Protects and Prolongs Renal Graft Function During Cold Ischemia/Reperfusion and After Renal Transplantation

Since the discovery of the therapeutic potential of H<sub>2</sub>S, there has been more focus on the protective role of H<sub>2</sub>S in warm IRI in the kidney compared with cold IRI in renal

transplantation. However, emerging evidence suggests that exogenous H<sub>2</sub>S administration protects renal cells against cold I/R-induced apoptosis, vasoconstriction, oxidative stress, and inflammation and thus increases the survival rate of the renal graft after transplantation. For example, following 18 h of cold (4°C) storage, NaHS treatment at 10 min before and after reperfusion has been shown to enhance renal blood flow (RBF) and increase renal graft function of porcine kidneys (41). Studies by our own group have demonstrated that addition of NaHS to UW solution during preservation of rat donor kidneys significantly mitigates cold IRI and improves post-transplant renal graft function and survival following both moderate and severe periods of cold (4°C) storage (53, 55). Thus, H<sub>2</sub>S protects and prolongs renal graft function during cold I/R and after renal transplantation.

#### H<sub>2</sub>S scavenges ROS and preserves renal function during cold I/R

It is well known that ROS is a chief mediator of cold IRI in human donor kidneys and therefore the use of H<sub>2</sub>S as an antioxidant has been suggested to mitigate oxidative stress during cold storage and reperfusion. Hosgood and Nicholson (41) observed a significant reduction of oxidative stress in porcine renal graft following treatment with NaHS before and during reperfusion. We also found that dopamine treatment preserved kidney function and integrity *via* maintenance (and upregulation) of renal CBS, CSE, and 3-MST and elevation of serum H<sub>2</sub>S, resulting in a marked decrease in the levels of renal malondialdehyde (MDA; a product of lipid peroxidation in the cell and a marker of oxidative stress) in a rat model closely matching clinical whole-body deep cooling and re-warming (12). Moreover, pharmacological inhibition of H<sub>2</sub>S

production with amino-oxyacetic acid (AOAA) in dopamine-treated hypothermic rats strongly reduced renal expression of CBS, CSE, and 3-MST and serum H<sub>2</sub>S and significantly increased renal oxidative stress, resulting in severe renal injury (30).

This finding supports our previous study in which we showed that dopamine and serotonin doubled CBS expression *via* mammalian target of rapamycin, leading to increased endogenous H<sub>2</sub>S production and reduced ROS levels in cultured smooth muscle cells (81). This might also be the mechanism by which dopamine upregulates renal CSE and 3-MST under hypothermia/rewarming condition. Along the same train of thought, AOAA treatment also markedly reduced the serum H<sub>2</sub>S level (Fig. 3B) and creatinine clearance (Fig. 4B) in Syrian hamsters subjected to 5'-AMP-induced torpor-like state at 7°C body temperature (Fig. 3A) (29). However, a major caveat in the use of AOAA (and other endogenous H<sub>2</sub>S inhibitors) is its unspecificity to H<sub>2</sub>S-producing enzymes and therefore would require knockout/knockdown models to complement its use.

The antioxidant effect of H<sub>2</sub>S is also observed through therapeutic concentrations of the novel mitochondrial-targeting molecule, AP39 (49), which significantly reduced ROS production and preserved mitochondrial membrane integrity in our novel *in vitro* model of cold hypoxia/hypercapnia and warm reoxygenation (H/R) injury (54) as well as in other *in vitro* models of hypoxia/oxidative stress injury in renal epithelial cells and in a rat model of renal IRI (1), showing a direct mitochondrial effect of AP39. Mechanistically, H<sub>2</sub>S regulates ROS production by enhancing the transport of homocysteine, which is reduced to cysteine, serving as a substrate for the synthesis of glutathione (the most abundant antioxidant in cells) and inhibiting the activity of nicotinamide adenine dinucleotide phosphate oxidase (NADPH oxidase [NOX]; a major source of ROS in the kidney) (73). Although the effect of H<sub>2</sub>S on NOX expression and activity has not been described extensively in the context of renal transplantation, Han *et al.* (39) recently reported that H<sub>2</sub>S administration *via* NaHS increased total glutathione and other antioxidant levels and significantly decreased the expression of NOX4 and NOX1 (members of the NOX family), MDA production, and glutathione-to-oxidized glutathione ratio in a mouse model of warm renal IRI and in human vascular smooth muscle cells (62).

It is important to note that apart from functioning as an ROS scavenger, H<sub>2</sub>S also boosts the endogenous antioxidant defense system by activating other antioxidant enzymes, such as glutathione, catalase, superoxide dismutase, and nuclear factor erythroid 2-related factor 2 (Nrf-2), and by increasing levels of N-acetylcysteine (18, 75) (Fig. 5). However, considering the low concentration of endogenous H<sub>2</sub>S in blood and tissues, its ability to directly scavenge ROS and its direct interaction with oxidants cannot completely account for its antioxidant effect. Studies suggest that H<sub>2</sub>S interaction with nitric oxide and carbon monoxide (members of the gasotransmitter family) in the mitochondrial respiratory chain could activate their individual antioxidant activities *via* activation and opening of ATP-sensitive potassium (K<sup>+</sup><sub>ATP</sub>) channels (61, 66, 94) and therefore could serve as a backup mechanism of the antioxidant activities of H<sub>2</sub>S. In conclusion, H<sub>2</sub>S treatment offers protection against cold IRI partly through its potent antioxidant action, which also includes recruitment and activation of other endogenous antioxidant pathways.

### *H<sub>2</sub>S maintains mitochondrial homeostasis and inhibits apoptosis during cold renal I/R*

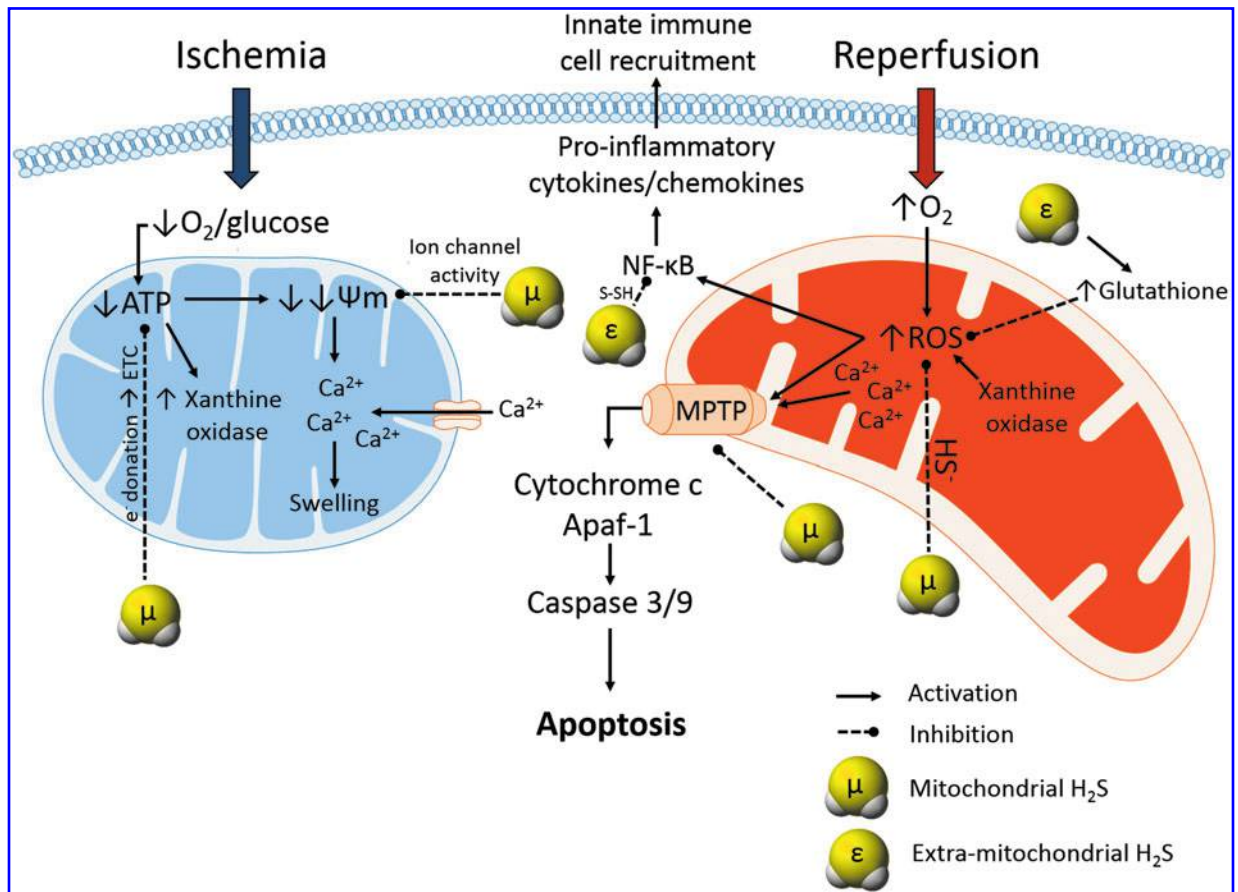
We have previously shown that H<sub>2</sub>S treatment protects donor kidneys against cold IRI at least partly by decreasing apoptosis in a rat model of renal transplantation. This was evidenced by a marked reduction in rates of renal graft apoptosis in conjunction with decreased expression of the proapoptotic gene, BH3-interacting domain death agonist (BID), and upregulation of the antiapoptotic gene, extracellular signal-regulated kinase-1 (ERK-1), compared with UW-treated renal grafts without H<sub>2</sub>S supplementation (53, 55). Similar to our studies, a recent study by Meng *et al.* demonstrated that the antiapoptotic role of H<sub>2</sub>S was associated with an increased expression of the antiapoptotic protein, Bcl-2, and decreased expression and activities of proapoptotic proteins (p38, Bax, and caspase-3) in both *in vitro* and *in vivo* models (58).

Considering both the increase in systemic H<sub>2</sub>S levels and the importance of mitochondrial protection for preservation of renal function during mammalian hibernation, it has been increasingly suggested that H<sub>2</sub>S may preserve mitochondrial integrity in a similar manner during cold IRI. We have recently produced novel and exciting evidence that supports this potential protective mechanism of H<sub>2</sub>S. In our study, we utilized AP39 to investigate whether targeting H<sub>2</sub>S release to mitochondria improves the potency of its protective effects during cold renal IRI. Using an *in vitro* model of H/R injury, we demonstrated that treatment of rat kidney cells with nanomolar concentrations of AP39 was at least 1000-fold more potent at preserving cellular viability following cold H/R injury compared with similar concentrations of the nonspecific H<sub>2</sub>S donor, GYY 4137 (54). Treatment of renal grafts with AP39 during prolonged cold storage also resulted in improved post-transplant graft function and survival, suggesting that targeting of H<sub>2</sub>S delivery to mitochondria could represent a viable and potent clinical strategy for minimizing the effect of IRI during transplantation (54).

Furthermore, AP39 stimulates mitochondrial respiration and generates mitochondrial sulfide and persulfide (49, 86), all of which act to maintain mitochondrial function during periods of hypoxia and thus make it an ideal candidate for organ preservation. Additionally, H<sub>2</sub>S (NaHS and AP39) inhibited Ca<sup>2+</sup>-mediated opening of mitochondrial permeability transition pore (MPTP) *via* a cyclophilin D-independent mechanism, preventing the release of proapoptotic factors from the mitochondria of hypertensive rat heart and also protecting rat myocardium against IRI (24, 44, 77). Hence, as hypothermia stimulates Ca<sup>2+</sup> influx and mitochondrial Ca<sup>2+</sup> accumulation (16), it is highly possible that H<sub>2</sub>S treatment of donor kidneys blocks Ca<sup>2+</sup> influx and mitochondrial Ca<sup>2+</sup> accumulation during cold storage by exerting a direct inhibitory effect on MPTP at a different site than cyclophilin D, thereby preventing the opening of MPTPs and preserving mitochondrial integrity (Fig. 5). Taken together, exogenous H<sub>2</sub>S appears to exert an antiapoptotic effect during cold renal IRI, which could be mediated by inhibitory actions against MPTP formation.

### *H<sub>2</sub>S induces renal vasodilation during cold I/R*

Cold storage of donor organs induces vasoconstriction, which has been partially implicated in cold-induced organ injury during transplantation. Emerging reports indicate that H<sub>2</sub>S induces vasodilation and thereby mitigates injurious



**FIG. 5. Proposed mechanisms of H<sub>2</sub>S-mediated protection of mitochondria during cold IRI.** During ischemia, ATP depletion causes inhibition of mitochondrial Na<sup>+</sup>/K<sup>+</sup> ion channels, resulting in decreased mitochondrial membrane potential (Ψ<sub>m</sub>), influx of Ca<sup>2+</sup> ions, and subsequent swelling of mitochondria. Prolonged periods of ischemia can permanently damage complexes in the ETC, which produce a burst of ROS as oxygen floods cells upon reperfusion. ROS are also produced in mitochondria by conversion of xanthine dehydrogenase to xanthine oxidase during ischemia. Xanthine oxidase converts hypoxanthine to uric acid, producing superoxide as a by-product. High levels of mitochondrial ROS production combined with Ca<sup>2+</sup>-induced mitochondrial swelling result in severe damage of mitochondrial membranes and formation of MPTPs, releasing proapoptotic factors, cytochrome c and apoptosis-inducing factor-1 (Apaf-1), which then activate the caspase 9/3 apoptotic signaling cascade, initiating cellular apoptosis. ROS-mediated mitochondrial damage also results in activation of proinflammatory NF-κB signaling and ultimately innate immune cell recruitment. H<sub>2</sub>S is proposed to directly mitigate the following mitochondrial mechanisms during IRI pathogenesis: (i) prevent loss of Ψ<sub>m</sub> and subsequent Ca<sup>2+</sup> influx through stimulation of ATP production via donation of electrons to the ETC and modulation of Ca<sup>2+</sup> channel activity; (ii) inhibit ROS production through direct inhibition (in HS<sup>-</sup> form); and (iii) directly inhibit MPTP formation via a cyclophilin D-independent mechanism. These mechanisms could be modulated through administration of external mitochondrial-targeted sources of H<sub>2</sub>S, such as AP39. Additionally, external sources of extramitochondrial H<sub>2</sub>S could indirectly blunt mitochondrial mechanisms of IRI pathogenesis via upregulation of the antioxidant glutathione and modulate proinflammatory NF-κB signaling via S-sulphydration of the p65 subunit. The actions of mitochondrial (μ) and extra-mitochondrial (ε) sources of H<sub>2</sub>S are, respectively, indicated in the figure above. ATP, adenosine triphosphate; IRI, ischemia/reperfusion injury; NF-κB, nuclear factor-kappa B; ROS, reactive oxygen species.

effects of cold-induced vasoconstriction. Xia *et al.* (89) first demonstrated that infusion of NaHS in intrarenal arteries in rats increases RBF and glomerular filtration rate (GFR) by increasing vasodilation in pre- and postglomerular arterioles and decreasing intrarenal vascular resistance, which would attenuate renal damage and improve oxygen and nutrient delivery to maintain functioning renal cells. This enhancement of RBF by H<sub>2</sub>S may result from activation and opening of K<sup>+</sup><sub>ATP</sub> channels and dilatation of blood vessels as this appears to be the underlying mechanism of protection by H<sub>2</sub>S in myocardial ischemia (93).

Following this initial demonstration of the effects of H<sub>2</sub>S on renal hemodynamics, Hosgood and Nicholson used a porcine model of renal transplantation to demonstrate that H<sub>2</sub>S treatment significantly increases RBF in donor kidneys 10 min before and after reperfusion compared with the control group without H<sub>2</sub>S supplementation (41). We have also observed that NaHS administration improves RBF and reduces the renal resistive index in a representative porcine model of donation after cardiac death (DCD) kidney transplantation (2). In addition, we subsequently developed a rat model of DCD renal transplantation to show that treatment of

DCD kidneys with H<sub>2</sub>S during both warm and cold IRIs results in decreased post-transplant graft injury and improved subsequent graft function and survival (37). Thus, by improving resistive index and RBF, H<sub>2</sub>S may improve perfusion parameters, which then may translate to improved renal function upon transplantation (Fig. 6).

It is important to point out that cyclosporine, a widely used immunosuppressive drug after renal transplantation, induces nephrotoxicity, which begins with renal and systemic vasoconstriction, leading to reduced GFR and ultimately interstitial fibrosis and tubular atrophy (72). Interestingly, a recent study has shown that H<sub>2</sub>S infusion before and after reperfusion of cold-stored porcine kidneys reverses cyclosporine-induced vasoconstriction and glomerular atrophy by inducing vasodilation and increasing RBF, suggesting an additional protective role of H<sub>2</sub>S-induced vasodilation during the transplant procedure (50). Thus, the vasodilatory effects of

H<sub>2</sub>S appear to reduce renal resistance and increase RBF in donor kidneys following cold storage, resulting in improved reperfusion and function of renal grafts in the acute post-transplant period.

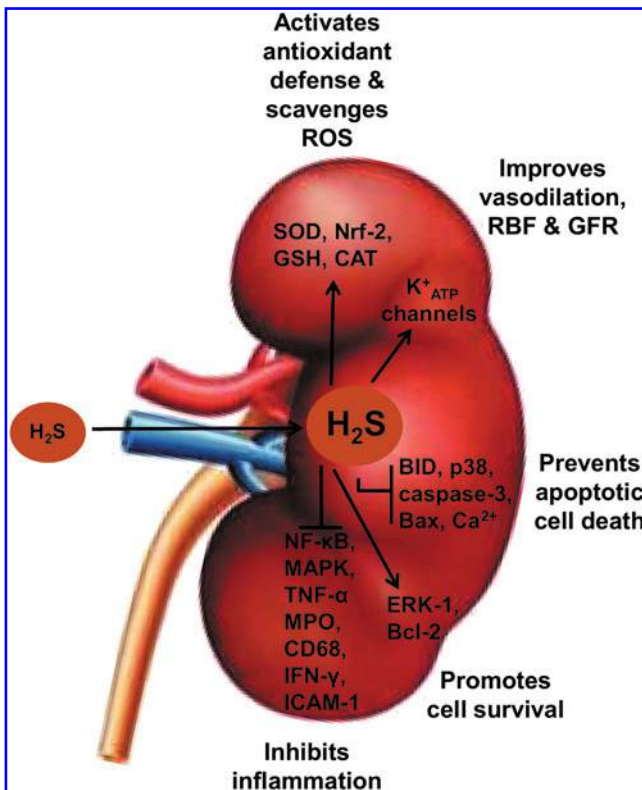
#### *H<sub>2</sub>S suppresses renal inflammation induced by cold I/R*

Apart from antioxidant, antiapoptotic, and vasodilatory effects, the therapeutic properties of H<sub>2</sub>S in renal transplantation also include an anti-inflammatory component. For example, we have previously demonstrated that H<sub>2</sub>S treatment of rat renal graft during cold storage significantly decreased infiltration of myeloperoxidase (MPO)-positive neutrophils and CD68-positive macrophages and expression of the proinflammatory cytokines interferon-gamma (IFN- $\gamma$ ) and intercellular adhesion molecule-1 (ICAM-1) in the acute post-transplant period (55) (Fig. 5). Our finding was confirmed by a recent study in which H<sub>2</sub>S administration suppressed influx of MPO-positive neutrophils in the kidneys of rats subjected to warm IRI (1).

While the anti-inflammatory property of H<sub>2</sub>S in renal transplantation remains to be revealed, other studies have reported more potent anti-inflammatory effects and mechanisms of H<sub>2</sub>S in various disease models (58). The slow-releasing H<sub>2</sub>S donor, GYY4137, inhibits nuclear factor-kappa B (NF- $\kappa$ B) and mitogen-activated protein kinases (p38, ERK1/2, and JNK1/2) (88), two key mediators of inflammation. Such anti-inflammatory effect of H<sub>2</sub>S was partially involved in myocardial protection through increased plasma H<sub>2</sub>S concentration and CSE activity in a rat model of myocardial IRI (58). H<sub>2</sub>S inhibits the expression of leukocyte adhesion molecules, thereby preventing leukocyte rolling and attachment to the endothelium (91). This inhibition was associated with activation of K<sup>+</sup><sub>ATP</sub> channels by H<sub>2</sub>S as K<sup>+</sup><sub>ATP</sub> antagonist reversed the leukocyte inhibitory effect of H<sub>2</sub>S and increased leukocyte adherence (91). These mechanisms underlying the anti-inflammatory action of H<sub>2</sub>S may thus partly account for renal protection against cold IRI in transplantation.

#### **Protective Role of H<sub>2</sub>S Against Cold IRI During Transplantation of Other Solid Organs**

Exogenous H<sub>2</sub>S treatment also proved protective against cold IRI in transplantation of various solid organs other than the kidney. In a rabbit model of lung transplantation, studies by George *et al.* showed that inhalation of gaseous H<sub>2</sub>S at 150 ppm for 2 h before 18 h of cold storage significantly decreased the ROS level, improved oxygenation, lowered pulmonary artery pressure, and prevented cold ischemic injury, thus leading to improved graft function during reperfusion compared with the control group (33, 34). This suggests that H<sub>2</sub>S is a potent scavenger of free radicals as discussed above. In a different experimental setting of cold IRI during lung transplantation, 3 h of cold storage, followed by NaHS treatment 15 min before reperfusion, significantly increased lung tissue expression of CSE and H<sub>2</sub>S levels and reduced membrane lipid oxidation levels, resulting in improved pulmonary function and lung protection following transplantation compared with control rats (87). Administration of propargylglycine (PPG; CSE inhibitor) 15 min before reperfusion markedly decreased lung CSE protein and H<sub>2</sub>S levels and increased lipid peroxidation and MPO activity,



**FIG. 6. Effects and mechanisms of H<sub>2</sub>S against cold IRI in renal transplantation.** Exogenous administration of H<sub>2</sub>S augments the endogenous H<sub>2</sub>S level, thereby stimulating the opening of K<sup>+</sup><sub>ATP</sub> channels, causing vasodilation and consequently increasing RBF and GFR. H<sub>2</sub>S also inhibits the activation of apoptotic and inflammatory pathways and up-regulates antiapoptotic genes such as ERK-1 and Bcl-2 and thus promotes renal cell survival. Additionally, H<sub>2</sub>S scavenges ROS and also activates endogenous antioxidants such as SOD, GSH, CAT, and Nrf-2, thereby bolstering the endogenous antioxidant defense system and thus reducing mitochondrial oxidative stress. Bcl-2, B cell lymphoma-2; CAT, catalase; ERK-1, extracellular signal-regulated kinase-1; GFR, glomerular filtration rate; GSH, glutathione; K<sup>+</sup><sub>ATP</sub>, ATP-sensitive potassium; Nrf-2, nuclear factor erythroid 2-related factor 2; RBF, renal blood flow; SOD, superoxide dismutase.



culminating in overproduction of ROS and pulmonary injury after transplantation (87). This suggests that reduced production and malfunction of endogenous H<sub>2</sub>S contribute to cold IRI in the lung.

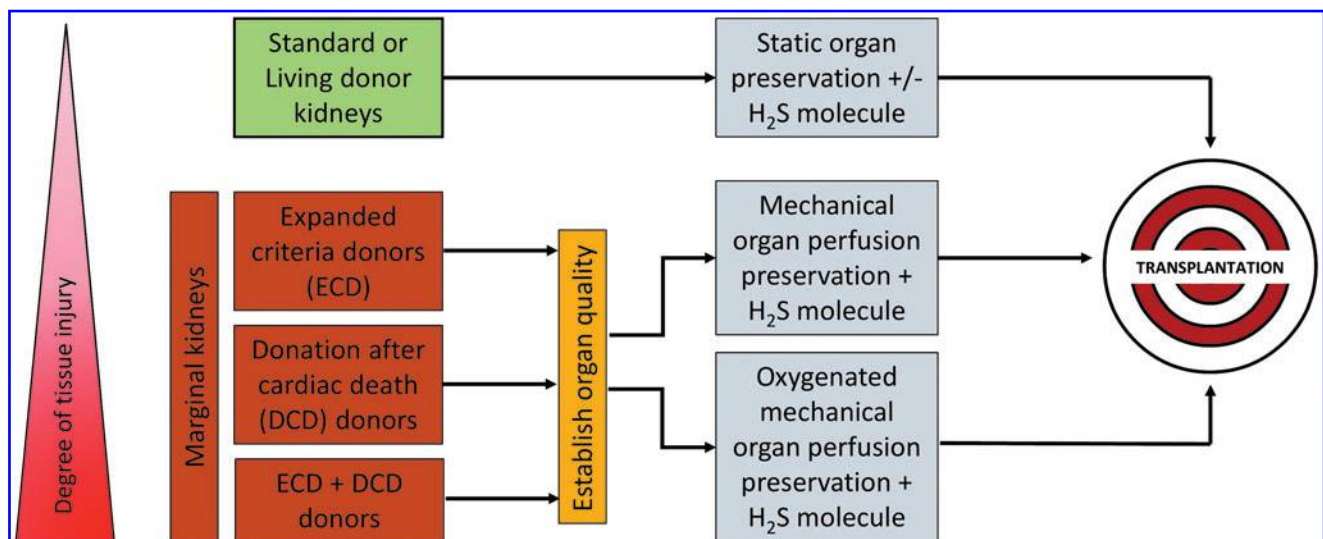
These findings also suggest that both endogenous and exogenous H<sub>2</sub>S treatments protect against cold IRI partly *via* their antioxidant properties. Although the mechanism by which PPG downregulates CSE expression and activity within such a short time is unknown, it is possible that lung CSE-producing cells are more sensitive to PPG than those of other organs, which might have resulted in death of CSE-producing cells within this short period. Moreover, the dose of PPG (37.5 mg/kg) used in the study might be higher than that used in other studies. This, therefore, needs further investigation. In a rat model of DCD liver transplantation, Na<sub>2</sub>S and gaseous H<sub>2</sub>S treatments increased the levels and activities of liver antioxidants and strongly reduced the levels of lactate dehydrogenase, MDA, and other liver damage markers during cold I/R and hence improved liver microcirculation, morphology, and function (3, 4). Thus, H<sub>2</sub>S-mediated cytoprotection against tissue IRI observed during renal transplantation is translatable to other tissues and should be further investigated to improve the outcomes of all solid organ transplantations.

### Conclusion and Future Perspectives

To our knowledge, there are no therapeutic strategies that can successfully mitigate IRI associated with organ transplantation. Therefore, the therapeutic use of exogenous H<sub>2</sub>S added to standard preservation solutions or inducing endogenous H<sub>2</sub>S seems a promising approach to prolong renal graft preservation and mitigate unavoidable cold IRI during transplantation of kidney and other solid organs (Fig. 7). Targeting exogenous

H<sub>2</sub>S administration to mitochondria increases the potency of its protective effects during IRI, implicating mitochondria as a primary site of action for H<sub>2</sub>S-mediated renoprotection. Exogenous H<sub>2</sub>S could potentially control mitochondrial mechanisms of IRI pathogenesis directly by driving ATP synthesis *via* donation of electrons to the ETC (36, 60), modulation of mitochondrial Ca<sup>2+</sup> channels to prevent Ca<sup>2+</sup> influx (82, 85), direct inhibition of mitochondrial ROS (20, 63), and prevention of MPTP formation (77) or indirectly *via* inhibition of proinflammatory NF-κB signaling (71) and upregulation of cellular antioxidant enzymes such as glutathione (46, 90).

Although extensive investigation in defining molecular mechanisms behind IRI and the use of various therapeutic strategies have been proposed, ongoing clinical studies, including those utilizing rPSGL-Ig (a p-selectin inhibitor) and mitoQ (an antioxidant), have provided some satisfying results in preventing renal IRI (25). In all cases, more clinical trials with improved standardization of protocols are required to contribute to important advances in clinical therapy (65). Apart from involvement of H<sub>2</sub>S in transplantation, additional lessons can be learned from dissecting mechanisms by which nature protects organs of hibernating animals from the damaging effects of cold exposure and reperfusion. Understanding such mechanisms will be beneficial for the development of more potent forms of the current therapeutic or novel therapeutic agents. In addition, future investigations should be aimed at developing new drugs or a combination of current therapeutic agents that could afford a multitarget approach to minimize IRI during transplantation. In this way, we may be able to take advantage of natural cytoprotective mechanisms utilized by hibernating mammals, such as H<sub>2</sub>S-mediated mitochondrial preservation, to ultimately improve clinical outcomes and quality of life for renal transplant recipients.



**FIG. 7. Potential model of kidney preservation before transplantation.** As donor organs become more marginal (ECD: age >60 or age >50 plus two of the following features—hypertension, terminal serum creatinine >1.5 mg/dl, or death from cerebrovascular accident; DCD donors), better evaluative tools will need to be devised to better establish organ quality before transplantation. Once this has been carried out, transplant teams may decide whether or not the organ should be preserved using static or mechanical organ preservation in standard solutions. There is strong experimental evidence to suggest that supplementing these standard solutions with H<sub>2</sub>S donor molecules may have a significant impact on both short- and long-term graft outcomes by modulating the deleterious effects of ischemia/reperfusion injury. DCD, donation after cardiac death; ECD, expanded criteria donors.

## References

- Ahmad A, Olah G, Szczesny B, Wood ME, Whiteman M, and Szabo C. AP39, A mitochondrially targeted hydrogen sulfide donor, exerts protective effects in renal epithelial cells subjected to oxidative stress in vitro and in acute renal injury in vivo. *Shock* 45: 88–97, 2016.
- Alabbassi A, Tran KC, Bloch M, Deng J, Mok A, Liu W, Garcia B, Sener A, and Luke P. The effects of molecules CORM-3 and H<sub>2</sub>S on renal protection during pulsatile perfusion. *Am J Transplant* 11: 500, 2011.
- Balaban CL, Rodriguez JV, and Guibert EE. Delivery of the bioactive gas hydrogen sulfide during cold preservation of rat liver: Effects on hepatic function in an *ex vivo* model. *Artif Organs* 35: 508–515, 2011.
- Balaban CL, Rodriguez JV, Tiribelli C, and Guibert EE. The effect of a hydrogen sulfide releasing molecule (Na<sub>2</sub>S) on the cold storage of livers from cardiac dead donor rats. A study in an *ex vivo* model. *Cryobiology* 71: 24–32, 2015.
- Barnes BM. Freeze avoidance in a mammal: Body temperatures below 0 degree C in an Arctic hibernator. *Science* 244: 1593–1595, 1989.
- Baumgart K, Radermacher P, and Wagner F. Applying gases for microcirculatory and cellular oxygenation in sepsis: Effects of nitric oxide, carbon monoxide, and hydrogen sulfide. *Curr Opin Anesthesiol* 22: 168–176, 2009.
- Beltowski J. Hypoxia in the renal medulla: Implication for hydrogen sulfide signaling. *J Pharmacol Exp Ther* 334: 358–363, 2010.
- Blackstone E, Morrison M, and Roth MB. H<sub>2</sub>S induces a suspended animation-like state in mice. *Science* 308: 518, 2005.
- Blackstone E and Roth MB. Suspended animation-like state protects mice from lethal hypoxia. *Shock* 27: 370–372, 2007.
- Bon D, Chatauret N, Giraud S, Thuillier R, Favreau F, and Hauet T. New strategies to optimize kidney recovery and preservation in transplantation. *Nat Rev Nephrol* 8: 339–347, 2012.
- Bos EM, Leuvenink HG, Snijder PM, Kloosterhuis NJ, Hillebrands JL, Leemans JC, Florquin S, and van Goor H. Hydrogen sulfide-induced hypometabolism prevents renal ischemia/reperfusion injury. *J Am Soc Nephrol* 20: 1901–1905, 2009.
- Bouma HR, Dugbartey GJ, Boerema AS, Talaei F, Herwig A, Goris M, van Buiten A, Strijkstra AM, Carey HV, Henning RH, and Kroese FGM. Reduction of body temperature governs neutrophil retention in hibernating and non-hibernating animals by margination. *J Leukoc Biol* 94: 431–437, 2013.
- Bouma HR, Kroese FG, Kok JW, Talaei F, Boerema AS, Herwig A, Draghiciu O, van Buiten A, Epema AH, van Dam A, Strijkstra AM, and Henning RH. Low body temperature governs the decline of circulating lymphocytes during hibernation through sphingosine-1-phosphate. *Proc Natl Acad Sci U S A* 108: 2052–2057, 2011.
- Bouma HR, Verhaag EM, Otis JP, Heldmaier G, Swoap SJ, Strijkstra AM, Henning RH, and Carey HV. Induction of torpor: Mimicking natural metabolic suppression for biomedical applications. *J Cell Physiol* 227: 1285–1290, 2012.
- Braunwald E and Bonow RO. *Braunwald's Heart Disease: A Textbook of Cardiovascular Medicine*. Philadelphia, PA: Saunders, 2012, p. xxiv.
- Brinkkoetter PT, Song H, Lösel R, Schnetzke U, Gottmann U, Feng Y, Hanusch C, Beck GC, Schnuelle P, Wehling M, van der Woude FJ, and Yard BA. Hypothermic injury: The mitochondrial calcium, ATP and ROS love-hate triangle out of balance. *Cell Physiol Biochem* 22: 195–204, 2008.
- Brown DC, Mulhausen RO, Andrew DJ, and Seal US. Renal function in anesthetized dormant and active bears. *Am J Physiol* 220: 293–298, 1971.
- Calvert JW, Jha S, Gundewar S, Elrod JW, Ramachandran A, Pattillo CB, Kevil CG, and Lefer DJ. Hydrogen sulfide mediates cardioprotection through Nrf2 signaling. *Circ Res* 105: 365–374, 2009.
- Camici PG, Prasad SK, and Rimoldi OE. Stunning, hibernation and assessment of myocardial viability. *Circulation* 117: 103–114, 2008.
- Carballal S, Trujillo M, Cuevasanta E, Bartesaghi S, Möller MN, Folkes LK, Garcia-Bereguian MA, Gutiérrez-Merino C, Wardman P, Denicola A, Radi R, and Alvarez B. Reactivity of hydrogen sulfide with peroxyxynitrite and other oxidants of biological interest. *Free Radic Biol Med* 50: 196–205, 2011.
- Carden DL and Granger DN. Pathophysiology of ischemia-reperfusion injury. *J Pathol* 190: 255–266, 2000.
- Carey HV, Andrews MT, and Martin SI. Mammalian hibernation: Cellular and molecular responses to depressed metabolism and low temperature. *Physiol Rev* 83: 1153–1181, 2003.
- Cavallo MC, Sepe V, Conte F, Abelli M, Ticozzelli E, Bottazzi A, and Geraci PM. Cost-effectiveness of kidney transplantation from DCD in Italy. *Transplant Proc* 46: 3289–3296, 2014.
- Chatzianastasiou A, Bibli SI, Andreadou I, Efentakis P, Kaludercic N, Wood ME, Whiteman M, Di Lisa F, Daiber A, Manolopoulos VG, Szabó C, and Papapetropoulos A. Cardioprotection by H<sub>2</sub>S donors: Nitric oxide-dependent and -independent mechanisms. *J Pharmacol Exp Ther* 358: 431–440, 2016.
- Dare AJ, Bolton EA, Pettigrew GJ, Bradley JA, Saeb-Parsy K, and Murphy MP. Protection against renal ischemia-reperfusion injury in vivo by the mitochondria targeted antioxidant MitoQ. *Redox Biol* 5: 163–168, 2015.
- de Vrij EL, Vogelaar PC, Goris M, Houwertjes MC, Herwig A, Dugbartey GJ, Boerema AS, Strijkstra AM, Bouma HR, and Henning RH. Platelet dynamics during natural and pharmacologically induced torpor and forced hypothermia. *PLoS One* 9: e93218, 2014.
- Debout A, Foucher Y, Trebern-Launay K, Legendre C, Kreis H, Mourad G, Garrigue V, Morelon E, Buron F, Rostaing L, Kamar N, Kessler M, Ladrière M, Poignas A, Bliidi A, Soullillou JP, Giral M, and Dantan E. Each additional hour of cold ischemic time significantly increases the risk of graft failure and mortality following renal transplantation. *Kidney Int* 87: 343–349, 2015.
- Drew KL, Osborne PG, Frerichs KU, Hu Y, Koren RE, Hellenbeck JM, and Rice ME. Ascorbate and glutathione regulation in hibernating ground squirrels. *Brain Res* 851: 1–8, 1999.
- Dugbartey GJ, Bouma HR, Strijkstra AM, Boerema AS, and Henning RH. Induction of a torpor-like state by 5'-AMP does not depend on H<sub>2</sub>S production. *PLoS One* 10: e0136113, 2015.
- Dugbartey GJ, Talaei F, Houwertjes MC, Goris M, Epema AH, Bouma HR, and Henning RH. Dopamine treatment attenuates acute kidney injury in a rat model of deep hypothermia and rewarming—The role of renal H<sub>2</sub>S-producing enzymes. *Eur J Pharmacol* 769: 225–233, 2015.

31. Fleck CC and Carey HV. Modulation of apoptotic pathways in intestinal mucosa during hibernation. *Am J Physiol Regul Integr Comp Physiol* 289: R586–R595, 2005.
32. Fu M, Zhang W, Wu L, Yang G, Li H, and Wang R. Hydrogen sulfide (H<sub>2</sub>S) metabolism in mitochondria and its regulatory role in energy production. *Proc Natl Acad Sci U S A* 109: 2943–2948, 2012.
33. George TJ, Arnaoutakis GJ, Beaty CA, Jandu SK, Santhanam L, Berkowitz DE, and Shah AS. Hydrogen sulfide decreases reactive oxygen in a model of lung transplantation. *J Surg Res* 178: 494–501, 2012.
34. George TJ, Arnaoutakis GJ, Beaty CA, Jandu SK, Santhanam L, Berkowitz DE, and Shah AS. Inhaled hydrogen sulfide improves graft function in an experimental model of lung transplantation. *J Surg Res* 178: 593–600, 2012.
35. Geró D, Torregrossa R, Perry A, Waters A, Le-Trionnaire S, Whatmore JL, Wood M, and Whiteman M. The novel mitochondria-targeted hydrogen sulfide (H<sub>2</sub>S) donors AP123 and AP39 protect against hyperglycemic injury in microvascular endothelial cells in vitro. *Pharmacol Res* 113: 186–198, 2016.
36. Gubern M, Andriamihaja M, Nubel T, Blachier F, and Bouillaud F. Sulfide, the first inorganic substrate for human cells. *FASEB J* 21: 1699–1706, 2007.
37. Grewal J, Lobb I, Saha M, Haig A, Jiang J, and Sener A. MP29-16 hydrogen sulfide supplementation mitigates effects of ischemia reperfusion injury in a murine model of donation after cardiac death renal transplantation. *J Urol* 195: e386, 2016.
38. Haak LL, Mignot E, Kilduff TS, Dement WC, and Heller HC. Regional changes in central monoamine and metabolic levels during the hibernation cycle in the golden-mantled ground squirrel. *Brain Res* 563: 215–220, 1991.
39. Han SJ, Kim JI, Park JW, and Park KM. Hydrogen sulfide accelerates the recovery of kidney tubules after renal ischemia/reperfusion injury. *Nephrol Dial Transplant* 30: 1497–1506, 2015.
40. Heldmaier G, Ortman S, and Elvert R. Natural hypometabolism during hibernation and daily torpor in mammals. *Respir Physiol Neurobiol* 141: 317–329, 2004.
41. Hosgood SA and Nicholson ML. Hydrogen sulphide ameliorates ischaemia-reperfusion injury in an experimental model of non-heart-beating donor kidney transplantation. *Br J Surg* 97: 202–209, 2010.
42. Jain S, Keys D, Martin S, Edelstein CL, and Jani A. Protection from apoptotic cell death during cold storage followed by rewarming in 13-lined ground squirrel tubular tells: The role of prosurvival factors X-linked inhibitor of apoptosis and phosphoAkt. *Transplantation* 100: 538–545, 2016.
43. Jani A, Epperson E, Martin J, Pacic A, Ljubanovic D, Martin SL, and Edelstein CL. Renal protection from prolonged cold ischemia and warm reperfusion in hibernating squirrels. *Transplantation* 92: 1215–1221, 2011.
44. Karwi QG, Bornbaum J, Boengler K, Torregrossa R, Whiteman M, Wood ME, Schulz R, and Baxter GF. AP39, a mitochondria-targeting hydrogen sulfide (H<sub>2</sub>S) donor, protects against myocardial reperfusion injury independently of salvage kinase signalling. *Br J Pharmacol* 174: 287–301, 2017.
45. Kashfi K and Olson KR. Biology and therapeutic potential of hydrogen sulfide and hydrogen sulfide-releasing chimeras. *Biochem Pharmacol* 85: 689–703, 2013.
46. Kimura Y, Goto Y, and Kimura H. Hydrogen sulfide increases glutathione production and suppresses oxidative stress in mitochondria. *Antioxid Redox Signal* 12: 1–13, 2010.
47. Kramarova LI, Lee TF, Cui Y, and Wang LC. State-dependent variation in the inhibitory effect of [D-Ala<sup>2</sup>, D-Leu<sup>5</sup>]-enkephalin on hippocampal serotonin release in ground squirrels. *Life Sci* 48: 175–181, 1991.
48. Lakhina AA, Markevich LN, Zakharova NM, Afanasyev VN, Kolomiytseva IK, and Fesenko EE. Phospholipids of liver cell nuclei during hibernation of Yakutian ground squirrel. *Dokl Biochem Biophys* 469: 235–238, 2016.
49. Le Trionnaire S, Perry A, Bartosz S, et al. The synthesis and functional evaluation of a mitochondria-targeted hydrogen sulfide donor, (10-oxo-10-(4-(3-thioxo-3H-1,2-dithiol-5-yl)-phenoxy)decyl)triphenyl phosphonium bromide (AP39). *Med Chem Commun* 5: 728–736, 2014.
50. Lee G, Hosgood SA, Patel MS, and Nicholson ML. Hydrogen sulphide as a novel therapy to ameliorate cyclosporine nephrotoxicity. *J Surg Res* 197: 419–426, 2015.
51. Li I, Whiteman M, Guan YY, et al. Characterization of a novel, water-soluble hydrogen sulfide-releasing molecule (GYY4137): New insights into the biology of hydrogen sulfide. *Circulation* 117: 2351–2360, 2008.
52. Li Q and Lancaster JR, Jr. Chemical foundations of hydrogen sulfide biology. *Nitric Oxide* 35: 21–34, 2013.
53. Lobb I, Davidson M, Carter D, Liu W, Haig A, Gunaratnam L, and Sener A. Hydrogen sulfide treatment mitigates renal allograft ischemia-reperfusion injury during cold storage and improves early transplant kidney function and survival following allogeneic renal transplantation. *J Urol* 194: 1806–1815, 2015.
54. Lobb I, Jiang J, Lian D, Liu W, Haig A, Saha MN, Torregrossa R, Wood ME, Whiteman M, and Sener A. Hydrogen sulfide protects renal grafts against prolonged cold ischemia-reperfusion injury via specific mitochondrial actions. *Am J Transplant* 17: 341–352, 2017.
55. Lobb I, Mok A, Lan Z, Liu W, Garcia B, and Sener A. Supplemental hydrogen sulfide protects transplant kidney function and prolongs recipient survival after prolonged cold ischemia-reperfusion injury by mitigating renal apoptosis and inflammation. *BJU Int* 110: E1187–E1195, 2012.
56. Ma YL, Zhu X, Rivera PM, Toien O, Barnes BM, LaManna JC, Smith MA, and Drew KL. Absence of cellular stress in brain after hypoxia induced by arousal from hibernation in Arctic ground squirrels. *Am J Physiol Regul Integr Comp Physiol* 289: R1297–R1306, 2005.
57. Malek M and Nematbakhsh M. Renal ischemia/reperfusion injury; from pathophysiology to treatment. *J Renal Inj Prev* 4: 20–27, 2015.
58. Meng G, Wang J, Xiao Y, Bai W, Xie L, Shan L, Moore PK, and Ji Y. GYY4137 protects against myocardial ischemia and reperfusion injury by attenuating oxidative stress and apoptosis in rats. *J Biomed Res* 29: 203–213, 2015.
59. Mitchell T, Saba H, Laakman J, Parajuli N, and Macmillan-Crow LA. Role of mitochondrial-derived oxidants in renal tubular cell cold-storage injury. *Free Radic Biol Med* 49: 1273–1282, 2010.
60. Modis K, Coletta C, Erdelyi K, Papapetropoulos A, and Szabo C. Intramitochondrial hydrogen sulfide production by 3-mecaptopyrivate sulfurtransferase maintains mitochondrial electron flow and supports cellular bioenergetics. *FASEB J* 27: 601–611, 2013.
61. Murphy ME and Brayden JE. Nitric oxide hyperpolarizes rabbit mesenteric arteries via ATP-sensitive potassium channels. *J Physiol* 486: 47–58, 1995.

62. Muzaffar S, Shukla N, Bond M, Newby AC, Angelini GD, Sparatore A, Del Soldato P, and Jeremy JY. Exogenous hydrogen sulfide inhibits superoxide formation, NOX-1 expression and Rac1 activity in human vascular smooth muscle cells. *J Vasc Res* 45: 521–528, 2008.
63. Nagy P and Winterbourn CC. Rapid reaction of hydrogen sulfide with the neutrophil oxidant hypochlorous acid to generate polysulfides. *Chem Res Toxicol* 23: 1541–1543, 2010.
64. Ojo AO, Wolfe RA, Held PJ, Port FK, and Schumouder RL. Delayed graft function: Risk factors and implications for renal allograft survival. *Transplantation* 63: 968, 1997.
65. Pantazi E, Bejoui M, Folch-Puy E, Adam R, and Rosello-Catafau J. Advances in treatment strategies for ischemia reperfusion injury. *Expert Opin Pharmacother* 17: 169–179, 2016.
66. Pereira de Avila MA, Giusti-Paiva A, and Giovani de Oliveira Nascimento C. The peripheral antinociceptive effect induced by the heme oxygenase/carbon monoxide pathway is associated with ATP-sensitive K<sup>+</sup> channels. *Eur J Pharmacol* 726: 41–48, 2014.
67. Quinones QJ, Ma Q, Zhang Z, Barnes BM, and Podgoreanu MV. Organ protective mechanisms common to extremes of physiology: A window through hibernation biology. *Integr Comp Biol* 54: 497–515, 2014.
68. Salahudeen AK, Haider N, and May W. Cold ischemia and reduced long-term survival of cadaveric renal allografts. *Kidney Int* 65: 713–718, 2004.
69. Sallmen T, Beckman AL, Stanton TL, et al. Major changes in the brain histamine system of the ground squirrel *Citellus lateralis* during hibernation. *J Neurosci* 19: 1824–1835, 1999.
70. Sandovici M, Henning RH, Hut RA, Strijkstra AM, Epema AH, van Goor H, and Deelman LE. Differential regulation of glomerular and interstitial endothelial nitric oxide synthase expression in the kidney of hibernating ground squirrel. *Nitric Oxide* 11: 194–200, 2004.
71. Sen N, Paul BD, Gadalla MM, Mustafa AK, Sen T, Xu R, Kim S, and Snyder SH. Hydrogen sulfide-linked sulfhydrylation of NF-kappaB mediates its antiapoptotic actions. *Mol Cell* 45: 13–24, 2012.
72. Senero J, Rodrigues-Santos P, Vala H, Rocha-Pereira P, Alves R, Fernandes J, Santos-Silva A, Carvalho E, Teixeira F, and Reis F. Transition from cyclosporine-induced renal dysfunction to nephrotoxicity in an in vivo rat model. *Int J Mol Sci* 15: 8979–8997, 2014.
73. Shan XQ, Aw TY, and Jones DP. Glutathione-dependent protection against oxidative injury. *Pharmacol Ther* 47: 61–71, 1990.
74. Shibuya N, Koike S, Tanaka M, Ishigami-Yuasa M, Kimura Y, Ogasawara Y, Fukui K, Nagahara N, and Kimura H. A novel pathway for the production of hydrogen sulfide from D-cysteine in mammalian cells. *Nat Commun* 4: 1366, 2013.
75. Shimada S, Fukai M, Wakayama K, Ishikawa T, Kobayashi N, Kimura T, Yamashita K, Kamiyama T, Shimamura T, Taketomi A, and Todo S. Hydrogen sulfide augments survival signals in warm ischemia and reperfusion of the mouse liver. *Surg Today* 45: 892–903, 2015.
76. Stenvinkel P, Jani AH, and Johnson RJ. Hibernating bears (Ursidae): Metabolic magicians of definite interest for the nephrologist. *Kidney Int* 83: 207–212, 2013.
77. Strutynska NA, Dorofeieva NO, Vavilova HL, and Sahach VF. Hydrogen sulfide inhibits Ca(2+)-induced mitochondrial transition pore opening in spontaneously hypertensive rats. *Fiziol Zh* 59: 3–10, 2013.
78. Stubenitsky BM, Booster MH, Kootstra G, Brasile L, and Haisch C. Deleterious effect of prolonged cold ischemia on renal function. *Transpl Int* 14: 256–260, 2001.
79. Szczesny B, Modis K, Yanagi K, Coletta C, Le Trionnaire S, Perry A, Wood ME, Whiteman M, and Szabo C. AP39, a novel mitochondria-targeted hydrogen sulfide donor, stimulates cellular bioenergetics, exerts cytoprotective effects and protects against the loss of mitochondrial DNA integrity in oxidatively stressed endothelial cells in vitro. *Nitric Oxide* 41: 120–130, 2014.
80. Talaei F, Bouma HR, Hylkema MN, Strijkstra AM, Boereman AS, Schmidt M, and Henning RH. The role of endogenous H<sub>2</sub>S formation in reversible remodeling of lung tissue during hibernation in the Syrian hamster. *J Exp Biol* 15: 2912–2919, 2012.
81. Talaei F, Bouma HR, van der Graaf AC, Strijkstra AM, Schmidt M, and Henning RH. Serotonin and dopamine protect from hypothermia/rewarming damage through the CBS/H<sub>2</sub>S pathway. *PLoS One* 6: e22568, 2011.
82. Tang G, Wu L, and Wang R. Interaction of hydrogen sulfide with ion channels. *Clin Exp Pharmacol Physiol* 37: 753–763, 2010.
83. Teng H, Wu B, Zhao K, Yang G, Wu L, and Wang R. Oxygen-sensitive mitochondria accumulation of cystathionine β synthase mediated by Lonprotease. *Proc Natl Acad Sci U S A* 110: 12679–12684, 2013.
84. Tøien O, Blake J, Edgar DM, Grahn DA, Heller HC, and Barnes BM. Hibernation in black bears: Independence of metabolic suppression from body temperature. *Science* 331: 906–909, 2011.
85. Tomasova L, Pavlovicova M, Malekova L, Misak A, Kristek F, Grman M, Cacanyiova S, Tomasek M, Tomaskova Z, Perry A, Wood ME, Lacinova L, Ondrias K, and Whiteman M. Effects of AP39, a novel triphenylphosphonium derivatised anethole dithiolethione hydrogen sulfide donor, on rat haemodynamic parameters and chloride and calcium Cav3 and RyR2 channels. *Nitric Oxide* 46: 131–144, 2015.
86. Wedmann R, Onderka C, Wei S, Szijártó IA, Miljkovic JL, Mitrovic A, Lange M, Savitsky S, Yadav PK, Torregrossa R, Harrer EG, Harrer T, Ishii I, Gollasch M, Wood ME, Galardon E, Xian M, Whiteman M, Banerjee R, and Filipovic MR. Improved tag-switch method reveals that thioredoxin acts as deperoxidase and controls the intracellular levels of protein persulfidation. *Chem Sci* 7: 3414–3426, 2016.
87. Wu J, Wei J, You X, Chen X, Zhu H, Zhu X, Liu Y, and Xu M. Inhibition of hydrogen sulfide generation contributes to lung injury after experimental orthotopic lung transplantation. *J Surg Res* 182: e25–e33, 2013.
88. Wu Z, Peng H, Lin W, and Liu Y. GYY4137, a hydrogen sulfide-releasing molecule, inhibits the inflammatory response by suppressing the activation of nuclear factor-kappa B and mitogen-activated protein kinases in Coxsackie virus B3-infected rat cardiomyocytes. *Mol Med Rep* 11: 1837–1844, 2015.
89. Xia M, Chen L, Muh RW, Li PL, and Li N. Production and actions of hydrogen sulfide, a novel gaseous bioactive substance, in the kidneys. *J Pharmacol Exp Ther* 329: 1056–1062, 2009.
90. Yang G, Zhao K, Ju Y, Mani S, Cao Q, Puukila S, Khaper N, Wu L, and Wang R. Hydrogen sulfide protects against cellular senescence via S-sulfhydrylation of

- Keap1 and activation of Nrf2. *Antioxid Redox Signal* 18: 1906–1919, 2013.
91. Zanardo RC, Brancaleone V, Distrutti E, Fiorucci S, Cirino G, and Wallace JL. Hydrogen sulfide is an endogenous modulator of leukocyte-mediated inflammation. *FASEB J* 20: 2118–2120, 2006.
  92. Zancanaro C, Malatesta M, Mannello F, Vogel P, and Fakan S. The kidney during hibernation and arousal from hibernation. A natural model of organ preservation during cold ischemia and reperfusion. *Nephrol Dial Transplant* 14: 1982–1990, 1999.
  93. Zhang Z, Huang H, Liu P, Tang C, and Wang J. Hydrogen sulfide contributes to cardioprotection during ischemia-reperfusion injury by opening K ATP channels. *Can J Physiol Pharmacol* 85: 1248–1253, 2007.
  94. Zhao W, Zhang J, Lu Y, and Wang R. The vasorelaxant effect of H(2)S as a novel endogenous gaseous K(ATP) channel opener. *EMBO J* 20: 6008–6016, 2001.

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#### Abbreviations Used

3-MST = 3-mercaptopyruvate sulfurtransferase  
 AMP = adenosine monophosphate  
 AOAA = amino-oxyacetic acid  
 ATP = adenosine triphosphate  
 Bcl-2 = B cell lymphoma-2  
 CBS = cystathionine  $\beta$ -synthase  
 CSE = cystathionine  $\gamma$ -lyase  
 DCD = donation after cardiac death  
 DGF = delayed graft function  
 ERK-1 = extracellular signal-regulated kinase-1  
 ETC = electron transport chain  
 GFR = glomerular filtration rate  
 H<sub>2</sub>S = hydrogen sulfide  
 H/R = hypoxia/hypercapnia and warm reoxygenation  
 I/R = ischemia/reperfusion  
 IRI = ischemia/reperfusion injury  
 K<sup>+</sup><sub>ATP</sub> = ATP-sensitive potassium  
 MDA = malondialdehyde  
 MPO = myeloperoxidase  
 MPTP = mitochondrial permeability transition pore  
 NADPH = nicotinamide adenine dinucleotide phosphate  
 NaHS = sodium hydrosulfide  
 NF- $\kappa$ B = nuclear factor-kappa B  
 NOX = NADPH oxidase  
 PPG = propargylglycine  
 RBF = renal blood flow  
 ROS = reactive oxygen species  
 UW = University of Wisconsin